1	Accelerating Silphium domestication: an opportunity to develop new crop ideotypes and
2	breeding strategies informed by multiple disciplines.
3	David L. Van Tassel,* Kenneth A. Albrecht, James D, Bever, Arvid A. Boe, Yaniv Brandvain,
4	Timothy E. Crews, Markus Gansberger, Pedro Gerstberger, Luciana González-Paleo, Brent S.
5	Hulke, Nolan C. Kane, Paul J. Johnson, Elena G. Pestsova, Valentín D. Picasso Risso, Jarrad R,
6	Prasifka, Damian A. Ravetta, Brandon Schlautman, Craig C. Sheaffer, Kevin P. Smith, Pablo R.
7	Speranza, M. Kathryn Turner, Alejandra E. Vilela, Philipp von Gehren, and Christian Wever.
8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26	David Van Tassel, Timothy Crews, Kathryn Turner and Brandon Schlautman, The Land Institute 2440 E Water Well Rd., Salina, KS 67448; Kenneth A. Albrecht and Valentín Picasso Risso, Department of Agronomy, University of Wisconsin-Madison, 1575 Linden Dr., Madison, WI; Arvid Boe, Plant Science, South Dakota State University, Box 2140C, Brookings, SD 57007; Yaniv Brandvain, Department of Plant Biology, University of Minnesota 232 Cargill Building 1500 Gortner Avenue St. Paul, MN 55108; Markus Gansberger and Philipp von Gehren, Austrian Agency for Health and Food Safety Ltd. (AGES) Spargelfeldstraße 191 1220 Wien, Austria; Pedro Gerstberger, Department of Plant Ecology, University of Bayreuth, Post box 10 12 51, D-95448 Bayreuth, Germany; Luciana González-Paleo, Damián Ravetta and Alejandra Vilela, National Scientific and Technical Research Council (MEF-CONICET), Fontana 140, Trelew, Chubut, Argentina; Brent Hulke and Jarrad Prasifka, Northern Crop Science Laboratory, USDA-ARS 1307 18th St. N. Fargo, ND 58102; Nolan Kane, Ecology and Evolutionary Biology, University of Colorado, 1900 Pleasant Street 334, UCB Boulder, CO 80309-0334; Paul Johnson, Insect Biodiversity Lab Box 2207A, Brookings, SD 57007; Craig Sheaffer and Kevin Smith, Department of Agronomy and Plant Genetics, University of Minnesota, 411 Borlaug Hall 1991 Upper Buford Circle, St. Paul, MN55108; Pablo Speranza, Faculty of Agronomy, University of the Republic, Garzón 780, 12900, Montevideo, Uruguay; Elena Pestsova and Christian Wever, Institute of developmental and molecular biology of plants, Heinrich-Heine-University University of Laboratory.

27 Received \_\_\_\_\_\_ \*Corresponding author (<u>dlvantassel@gmail.com</u>)

28

1 Abstract: Silphium perfoliatum L. (cup plant, silphie) and S. integrifolium Michx. (rosinweed, 2 silflower) are in the same sub-family and tribe as sunflower (*Helianthus annuus* L.). S. *perfoliatum* has been grown in many countries as a forage or bioenergy crop with forage quality 3 4 approaching that of alfalfa and biomass yield close to maize in some environments. S. integrifolium has large seeds with taste and oil quality similar to traditional oilseed 5 sunflower. Silphium species are all long-lived, diploid perennials. Crops from this genus could 6 improve the yield stability, soil, and biodiversity of agricultural landscapes because in their wild 7 state they are deep-rooted and support a wide diversity of pollinators. In contrast to pre-modern 8 domestication, *de novo* domestication should be intentional and scientific. We have the luxury 9 and obligation at this moment in history to expand the domestication ideotype from food/energy 10 production to include (1) crop-driven ecosystem services important for sustainability, (2) genetic 11 12 diversity to enable breeding progress for centuries, (3) natural adaptations and microbiome associations conferring resource use efficiency and stress tolerance, (4) improving domestication 13 theory itself by monitoring genetic and ecophysiological changes from pre-domestication 14 baselines. Achieving these goals rapidly will require the use of next generation sequencing for 15 marker development and an international, interdisciplinary team committed to collaboration and 16 strategic planning. 17

18

Abbreviations: G x E, genotype by environment; GWAS, genome wide association study; NGS,
next generation sequencing.

21

1 Pre-historic domestication proceeded by ongoing conscious and unconscious human selection, driven by the seed-dispersing behavior of humans attempting to secure short-term 2 survival (Diamond 2002). Thanks to them and modern plant breeders, there is currently 3 4 unprecedented production of grain which temporarily provides us the opportunity to critically 5 evaluate the results of past domestication and to plan for long-term human survival. Although our understanding of plant physiology, the plant microbiome, evolution and genetics, coupled 6 7 with the development of powerful new sequencing and gene editing techniques allows us 8 unprecedented opportunities to characterize our crops' gene-pools and accelerate ongoing crop improvement, today's food crops are committed to a narrow range of evolutionary trajectories. 9 For both biological reasons (polyploidy, extinct ancestors, narrowed gene pools, developmental 10 canalization) and cultural/economic reasons (commodity standards, farmer traditions, consumer 11 12 expectations) it would be very difficult to substantially reshape these crops to perform new ecosystem services or to thrive in very different environments (Juenger, 2013; Sharma et al., 13 2002; Rozema and Flowers, 2008). 14

If we could imagine an idealized case where humanity could start from scratch, and 15 16 conduct domestication from a scientific, long-term perspective, unconstrained by political pressure for rapid financial return on research investment, the needs of agricultural constituents, 17 or the burden of feeding today's hungry, what would we do differently? Would we want to 18 recreate crops functionally identical to our current major crops, or would the dramatically 19 different situation that humanity finds itself in now, compared with 10,000 years ago, require 20 new ideotypes or, at least, new expectations for the genetic structure and diversity of the crop 21 and its gene pools? Would we select from populations developed with time proven methods of 22 crossing 'good with good'? Alternatively, could more precise breeding enabled by modern 23

1 genomics allow for trait improvement with minimal linkage drag, and, consequently, minimal loss of genetic variation through selective sweeps and population bottlenecks? Perhaps breeders 2 may be able to select alleles with minimal undesirable pleiotropic effects? In the case of a closely 3 4 related species, sunflower, domestication was the result of the selective sweeping around many domestication loci, some very large, which put, perhaps unnecessarily, constraints on genetic 5 diversity of modern germplasm (Mandel et al., 2013; Baute et al., 2015). Therefore, well-planned 6 7 methods of domestication are warranted; general strategies for domestication and prioritization of candidates have been recently proposed (DeHaan et al., 2016). 8

As an example of a revision of the previous ideotypes for both new grain and biofuel 9 crops, the perennial habit has been proposed as a major improvement over the annual habit that 10 characterizes the majority of our current staple food and energy crops (Sanderson and Adler, 11 2008; Glover et al., 2010; Kantar et al., 2016). Breeding perennials for increased allocation to 12 13 sink structures such as stalks and seeds may reduce their fitness in unmanaged plant communities and reduce their longevity (González-Paleo and Ravetta, 2011) although DeHaan et al. (2007) 14 argued that a reduction from a lifespan of hundreds to tens of years might be acceptable. The 15 16 current deficit of high-yielding herbaceous perennial crops may be better explained by noting that species-rich, human-managed perennial crop fields represent a new environment to which no 17 existing species are adapted (Crews and DeHaan 2015), rather than invoking the impossibility of 18 plants employing two categorical adaptive strategies at the same time (Smaje, 2015). 19 Perennialism in plants is also something of a "gateway" character that opens the door to 20 botanical morphospace previously underutilized by agriculture (Van Tassel et al., 2010). If we 21 had the chance to start again, would we want to try to take advantage of some of the biochemical 22 pathways, leaf and root architectures, environmental adaptations, and defense mechanisms 23

1	contained in that untapped space? Perennialism also allows for agricultural exploitation of
2	desirable soil conditions, both physical and biological, that are more typically associated with
3	natural grassland or other wild ecosystems (Crews et al., 2016).
4	We have a unique opportunity to try <i>de novo</i> domestication as rationally as possible. Two
5	species of a North American prairie genus, Silphium, have been proposed as new crops. The
6	present authors have agreed to work together, sharing germplasm and expertise and coordinating
7	strategically on new experiments. We briefly review the features of Silphium that make it
8	suitable as a test case for "next generation domestication" and then outline our ideas of what that
9	means and how new approaches and tools may make it possible.

10

#### SILPHIUM AS A BLANK SLATE FOR DE NOVO DOMESTICATION 11

Silphium, a genus in the same tribe (Heliantheae) of the Asteraceae as Helianthus 12 13 (Clevinger and Panero, 2000), was noted by North American ecologists and conservationists in the 1930s and 40s for the ability of several of its species to tolerate many kinds of stress, 14 including the droughts of the Dust Bowl period (Weaver et al., 1935; Leopold, 1968). Silphium 15 species have been proposed for bioremediation of soils contaminated with crude oil or copper-16 mine tailings (Zhang et al., 2006, 2011) and revegetation of eroded, acidic landscapes in 17 Southern China (Ouyang et al., 2007). These reports suggest broad adaptation and tolerance to 18 general abiotic stresses. 19

**1. Economic botany.** The resinous sap characteristic of the genus (Bare, 1979) may have 20 been used as chewing gum by Native Americans and the roots as medicine (Stanford, 1990). 21

Although Leopold (1968) noted that cattle preferentially grazed the foliage and that the seeds
tasted like sunflower seeds, it was USSR scientists who first attempted to exploit the genus as a
forage crop beginning in 1957 (Stanford, 1990). Experimental plots of *S. perfoliatum* have
subsequently been grown in several European countries, China, Japan, Chile and the United
States since that time (Stanford, 1990; Gansberger et al., 2015), but large scale commercial use
has not been reported.

7 Experimental plots of S. integrifolium have been grown in Kansas and, independently in Poland since the early 2000s (Kowalski and Wiercinski, 2004; Van Tassel et al., 2014) and more 8 9 recently, plots have been established in Wisconsin, Minnesota, North Dakota, Texas, Vermont, Uruguay and Argentina. Whereas S. perfoliatum has always been considered as a potential 10 forage or bioenergy crop (Han et al., 2000a, 2000b; Lehmkuhler et al. 2007), S. integrifolium has 11 been considered as a potential new oilseed because its fatty acid profile is similar to the oilseed 12 13 sunflower (Kowalski and Wiercinski, 2004; Van Tassel et al., 2014). However, no commercial production has been initiated, reflecting the lack of improved cultivars and the many processing 14 and marketing difficulties faced by new crops (Raymer, 2002). 15

2. Genetics. All *Silphium* species are diploids, with 7 pairs of chromosomes, but have 16 roughly 2.5 times the genome size of sunflower (2C = 16.6 - 16.9 pg) (Bai et al., 2012, Table S1). 17 Molecular data has only been used for establishing the phylogenetic relationships within the 18 genus and sub-tribe (Clevinger and Panero, 2000) but otherwise both genomic organization and 19 20 the population genetic structure of these species remain unknown. Both S. perfoliatum and S. integrifolium cluster in the same sub-clade of the genus (Clevinger and Panero, 2000) and can be 21 crossed with normal pairing (Settle, 1967). Spontaneous hybrids are known (Yatskievych, 2006) 22 although such hybrids have reduced fertility (Van Tassel, unpublished data, 2016). It remains to 23

be seen if interspecific F<sub>1</sub> cultivars or stable allopolyploids could be viable options for combining
 the agronomic traits from different *Silphium* species. At the minimum, it seems likely that *S*.
 *perfoliatum* and *S. integrifolium* can be sources of alleles for each other using introgression
 breeding.

5 3. Plant Breeding. Exploratory genetic research has demonstrated heritable variation for traits of interest in both cases (Van Tassel et al., 2014; Assefa et al., 2015). Breeding 6 populations of S. *integrifolium* have been developed with increased numbers of seeds per head, 7 through selection for (pistillate) ray florets (Van Tassel, et al., 2014). Several improved 8 populations were developed in Russia and Ukraine in the past (Arkhipenko and Larina, 2011), 9 but no well-documented and widely available cultivars or genetic stocks have been released in 10 recent decades despite recent interest and expanding hectorage in Germany and farmer requests 11 for further improvement (Pestsova, personal communication, 2016, 2017). Seed dormancy and 12 13 seedling emergence and vigor are among the most limiting traits (Gansberger et al., 2015; Von Gehren et al., 2016). 14

15

## **16 OPPORTUNITIES TO DO DOMESTICATION DIFFERENTLY**

Opportunities to accelerate domestication using next-generation sequencing based techniques and insights from decades of advances in evolutionary biology have been reviewed elsewhere (Henry, 2012; Harfouche et al., 2012; Shapter et al., 2013; DeHaan and Van Tassel, 2014). If *Silphium* represents a new opportunity to do better than our ancient ancestors, we suggest that rational, intentional domestication requires us to consider not just why to use perennials, or how to do it faster, that is, how to efficiently increase yields and make the crop easier to harvest, use, and propagate, but how a newly domesticated crop might be intentionally
developed anticipating the stages of crop research and utilization following domestication. The
challenge of new perennial breeding programs will be to maintain those traits associated with the
perennial habit and the provision of ecosystem services while increasing yield potential and
stability (Vico et al., 2015).

6 1. Improved capacity for delivering specific ecosystem services. As the matrix of 7 native vegetation declines in agricultural regions, agroecologists are attempting to design agricultural landscapes where the crops themselves drive needed ecosystem processes (Power, 8 2010; Asbjornsen, 2013; Runck et al., 2014). While many wild perennial plants may help 9 stabilize soil and sequester carbon, the baseline allocation patterns, and the morphological and 10 functional traits that provide those and other types of services of domestication candidates should 11 be described in their pre-domesticated state. Understanding wild candidates in their ecosystem 12 13 context, including successional state and soil development, is also germane to the delivery of services, especially those associated with the soil microbiome (Koziol and Bever 2015). The 14 causal relationships between plant form and function and scalable services must be investigated 15 16 to help us track changes in key traits at the same time that we determine if they are compromised during domestication, and to do our best to ensure that they are not. Since not every trait can be 17 monitored, wild candidates should be assessed critically in order to prioritize the potential 18 services in which they likely have a comparative advantage over other crops. 19

*Silphium* species are known to be mycorrhizal (Dhillion and Friese, 1992), deep-rooted
(Wynia, 2009), drought tolerant (Weaver et al., 1935; Ouyang et al., 2007), and to support
populations of beneficial earthworms (Schorpp and Schrader, 2016) and a wide diversity of
insects (Fiedler and Landis, 2007; Tuell et al., 2008), including natural pest enemies (Kula et al.

1 2017). Deep-rooted perennial bioenergy crops can restore water quality through phytoextraction 2 of agrochemicals including nitrates (Ferchaud & Mary, 2016). Root architecture and mycorrhizal dependence can evolve rapidly as plants expand their range (Seifert et al., 2009). The potential 3 4 benefits of drought avoidance, nitrate scavenging, and mycorrhizal associations make these obvious traits to monitor during domestication. Other examples of relevant characters we are 5 looking at are those such as root, stem and leaf anatomy and composition that influence soil 6 organic matter formation. Changes in flowering phenology and pollen and nectar quantity and 7 quality should be tracked. *Silphium* secondary metabolites (Jamiolkowska and Kowalski, 2012) 8 are likely to influence its ecological impacts. Exactly which other services each Silphium crop 9 would be designed to deliver, and exactly which plant traits are required for fulfilling those 10 services, requires additional, ongoing research. Strategically chosen intercrops could provide 11 12 some services currently provided by wild *Silphium*. Polyculture could therefore reduce the complexity of the domesticated *Silphium* ideotype relating to services but complicate it by 13 adding requirements for interspecific facilitation or reduced competition. 14

We are committed to characterizing the wild plants and co-evolved symbionts, 15 16 mutualists, and community associates in order to more fully understand the strengths and weaknesses of these species with the goal of retaining traits that provide potential ecosystem 17 services and confer Silphium a comparative advantage over other crops. Wild and selected 18 candidates are being compared critically in Patagonia where early observations suggest that leaf 19 traits are already beginning to change as an unintended correlated response to selection for 20 reproductive traits (Vilela, unpublished data, 2016). Negative correlations between yield 21 components, classic domestication traits and ecosystem service traits will slow breeding progress 22 and need to be identified. 23

1	2. Considering the needs of future breeding efforts. The erosion of genetic diversity
2	during the domestication and modern improvement of major crops threatens to limit the ability of
3	breeders to further increase yields or adapt crops to new challenges. (Esquinas-Alcázar, 2005).
4	Although conversion of native grasslands to farmland has undoubtedly caused the extinction of
5	some local Silphium populations, nevertheless, S. integrifolium is currently found in 21 U.S.
6	states and S. perfoliatum in 31 states and 2 Canadian provinces (NRCS, 2016). Each, therefore,
7	can be found in a large geographical region and comprise several recognized sub-clades. This,
8	together with their longevity and strong self-incompatibility lead us to predict that they are
9	currently genetically diverse (J. Prasifka and B. Hulke, personal communication, 2016).
10	We plan to use three strategies to minimize the loss of genetic diversity during the
11	domestication of Silphium. First, both European and North American researchers have already
12	begun collecting seeds from wild populations throughout the range of both species in order to
13	greatly diversify the breeding populations (Pestsova, 2015; Wever, Brandvain, personal
14	communication, 2016). Use of the collected materials within a collaboration among Silphium
15	breeders and geneticists will allow for studies of Silphium genetic diversity and population
16	structure similar to those recently performed in domestication efforts for the potential bioenergy
17	crop Miscanthus (Clark et al., 2014, 2015), and ultimately, to ensure that Silphium domestication
18	begins with a broad base of genetically diverse germplasm from a wide geographic origin.
19	Identifying a core collection (Brown, 1989) and understanding the genetic and phenotypic
20	variation within it will help breeders prioritize crossing based on their needs at the
21	time. Expanding this collaboration to physiologists and ecologists, using the collection in
22	additional simultaneous studies of Silphium plant morphology and ecophysiology, and
23	categorizing the ecological and environmental conditions of the collections sites is assisting in

1	the development of ideotypes for Silphium and in predicting its potential areas of cultivation.
2	Additionally, sequencing and phenotyping samples from this population will allow for both
3	gene-environment associations (Rellstab et al., 2015), and genome-wide association studies
4	(Visscher et al., 2012) to identify loci of interest in a breeding program. Finally, a study of this
5	sort could also provide valuable insights about the locations of unique Silphium diversity
6	hotspots which are candidates for in situ conservation efforts (Pavek et al., 2003; Meilleur and
7	Hodgkin, 2004) or to strategically identify landscape positions or regions where novel variation
8	may reside and where additional germplasm collection efforts should be focused (Reyno et al.,
9	2012).
10	Second, frequent monitoring of the allelic richness of breeding populations can help
11	minimize allele extinction by drift or hitchhiking through targeted enrichment strategies. NGS
12	technologies are being used to generate genome-wide marker datasets for S. integrifolium

(Smith, unpublished data 2016) and *S. perfoliatum* (Pestsova, unpublished data, 2016) that can
assist in monitoring allele diversity and facilitate the use of large breeding populations
(Allendorf et al., 2010).

Finally, in anticipation of some unavoidable narrowing of the genetic base of *Silphium* cultigens, we are preserving wild seed collections. *Ex situ* seed conservation has limitations (Simmonds, 1962), so we will also establish field gene-banks, preferably in several locations, where the core collections will be allowed to undergo sexual reproduction (using controlled polycrossing to avoid intermating between accessions) every 3-5 years. It is our view that it will be impossible—and undesirable—to prevent some natural and human-mediated selection from taking place. Indeed, one of the goals of maintaining these collections is to allow and encourage Crop Sci. Accepted Paper, posted 04/04/2017. doi:10.2135/cropsci2016.10.0834

Page 12 of 32

adaptation to evolving pests, pollinator support, and changing climate. Traits that will facilitate
 the use of these materials in future breeding programs will be gradually improved by culling
 plants with severe disease symptoms, lodging, or sterility.

4

# 3. Breeding specialized cultivars to design profitable and sustainable cropping

5 systems. Hasty domestication could result in the loss of potentially useful "wild traits" 6 (Ladizinsky, 1985; Cowling et al., 2009) conferring biotic and abiotic stress tolerance or 7 specialized resource-use or life-history strategies. Even in recently domesticated species, such as the American cranberry, these wild traits were not monitored, and as a result, breeding activities 8 9 have compromised necessary plant anti-herbivory defenses in commercial cranberry varieties (Rodriguez-Saona et al., 2011). Likewise, Benrey et al. (1998) found that Brassica and 10 Phaseolus cultivated domesticates were more susceptible to phytophagous insects and 11 parasitoids than their uncultivated wild brethren. On the other hand, native perennials may be 12 13 surprisingly susceptible to native pests (Prasifka et al., 2012); Pest tolerance alleles may be rare until strong directional, artificial selection increases their frequency. Therefore, it would also be 14 hasty to reject candidate species or candidate functional traits after observing only a few 15 individuals from unselected populations. 16

Wild *Silphium* germplasm is currently adapted to multiple latitudes and temperature regimes, and has been noted to tolerate flooding (Stanford, 1990), wind and drought (Weaver et al., 1935). As expected, *Silphium* species are not as water-use efficient as C<sub>4</sub> grasses (Schoo et al., 2016), but they are likely to be more resource conservative than the ruderal ancestors of annual crops or perennial species strongly selected for rapid growth in resource rich environments (González-Paleo and Ravetta, 2011). The wild populations of both these *Silphium* species already show an ideotype of root architecture for drought resistance and nitrogen use efficiency, which was lost in several modern crops (Lynch 2013). It might therefore be possible
to domesticate both "acquisitive" and "conservative" biotypes of *Silphium* species which would
be better adapted to high and low input systems, respectively. Likewise, we plan to develop
breeding pools for several major geographic regions (cold temperate, sub-tropical, etc.). There is
a particular advantage of breeding *Silphium* species in cold regions since only a few C<sub>4</sub> species
can match the production of C<sub>3</sub> crops in cooler environments (Friesen et al. 2015)

In their native range, both *S. perfoliatum* and *S. integrifolium* show signs of being
unadapted to high-density monoculture, experiencing higher severity of insect herbivory and
fungal disease than in native plant communities (Johnson et al., 2012, Van Tassel et al.,
unpublished observations, 2016). In the short term, at least, breeding in and for "polyculture
adaptation" can prioritize yield potential and harvestability, while breeding for monoculture must
prioritize insect and disease tolerance or resistance before yield potential can be reliably
measured.

14 In an effort to maintain the broad genetic base and potential adaptability of Silphium through domestication, selection, and commercial cultivation, we propose to complement (A) 15 intensive selection for domestication traits, with (B) parallel programs to develop populations 16 with improved adaptation to new environments, and (C) heterotic groups. Approach B builds on 17 the long-term introgression strategy proposed by Simmonds (1993) by using population based 18 approaches to maximize recombination in large, genetically diverse and dynamic populations 19 20 that gradually become adapted to target environments through epistasis and coadaptation of genes (Haussmann et al., 2004). A third set of gene pools will be developed from the ex situ 21 22 collection (again, in genetic isolation from each other) in approach C to conserve genetic variation that contributes to heterosis. This variation could be lost in approach B, especially 23

where previously isolated populations are deliberately composited or used only as sources of
alleles for introgression into elite genetic backgrounds. After quantifying genetic distance using
modern methods to understand baseline population structure, targeted biparental crosses or
reciprocal recurrent selection among the *ex situ* accessions will be performed to analyze, enforce,
and enhance functional genetic divergence of heterotic pools (Saxena and Sawargaonkar, 2014;
Boeven et al., 2016).

New ideotypes being developed at this early stage of new domestication could also
include new options for achieving profitability. *Silphium* species have been found to contain a
range of phytochemicals, some with agricultural or industrial possibilities, including
antimicrobial activity (Jamiolkowska and Kowalski, 2012). Other products are being considered,
including resins (Ravetta, unpublished data, 2016), paper (Wever, personal communication,
2016) and solid fuel briquettes (Wrobel et al., 2013).

4. Providing data and theory for the domesticators of additional new crops. Like 13 14 most of evolution, the evolution of our present crops took place as unique, historical events. Much can be inferred from archaeological and comparative genetic/genomic studies, but much is 15 forever lost. With *de novo* domestication in the age of big data, gene editing and next generation 16 sequencing, we have the opportunity to more fully document the transition from wild plant to 17 crop. Because these are long-lived and can be propagated vegetatively (Vilela, unpublished data, 18 2016), some of the ancestral individuals from various stages of domestication will be cloned and 19 kept alive indefinitely enabling future scholars to replicate particular stages of the process. 20 Sequence and allele frequency data and dried tissue samples will also be archived. 21

22 Beyond archiving and documenting the process, we hope to conduct experimental

1 domestication involving replicated populations and selection regimes. The fact that two closely related species from the same genus are being domesticated simultaneously itself offers an 2 opportunity to compare their rate of morphological and genomic change and correlated responses 3 4 to selection. At least 2 sub-species of both S. perfoliatum and S. integrifolium are recognized ("The Plant List. Version 1.1.," 2013) and perhaps additional ecotypes or genetic clusters will be 5 identified; we plan to use the independent selection schemes in approaches A, B, and C above to 6 7 test hypotheses about the repeatability of genetic, physiological and anatomical changes that are thought to be associated with domestication (Gonzalez Paleo and Ravetta, 2012; Gonzalez Paleo 8 et al., 2016). Using both genetic and morphological data from breeding populations during each 9 generation of selection, we plan to perform selection signature studies to determine and compare 10 the genomic locations under artificial selection for the two species (Wright et al., 2010; 11 12 Prezeworski et al., 2005; Fontanesi et al., 2010). We imagine that such studies are of benefit not only to Silphium researchers, but also to a wider scientific audience interested in understanding 13 the genetic basis of adaptation and domestication. 14

15

### 16 NEXT GENERATION DOMESTICATION

Four general strategies recur when considering how to accomplish the goals described above.New technologies make each of these more feasible.

Multiple traits. Next-generation ideotypes are more complex than the previous
 generation, requiring the selection of traditional yield and domestication syndrome traits while
 avoiding loss of desirable wild characters. Multiple-trait genomic selection using multivariate
 models that take advantage of the genetic covariance between traits provides a practical means of

1 simultaneously selecting both wild and classical traits that make-up next-generation ideotypes (Jia and Jannink, 2012). Some of the "wild traits" or classical traits may be difficult or 2 expensive to measure, have low heritabilities, or may require too much time to measure. 3 4 However, generating genomic selection prediction models and/or using highly heritable indicator traits (genetically correlated with the wild or difficult to measure traits) will facilitate selection 5 progress without re-allocating substantial amounts of resources (Cameron, 1997; Calus and 6 7 Veerkamp, 2011). Moreover, multiple trait genomic selection models likely have special utility for breeding and domesticating perennial wild plants that have a more complex life-cycle and 8 morphology by improving selection accuracy and gain per unit time, which are generally limited 9 by periods of juvenility, and by allowing for selection of traits related to longevity (Heffner et 10 al., 2010; Resende et al., 2012; Desta and Ortiz, 2014; Gonzalez Paleo et al., 2016). On the other 11 12 hand, some necessary domestication traits may have high heritability and simple genetics, but are rare to discover in nature because they are unfit for natural environments. Monocephaly, non-13 dormant seed, or male sterility are examples. Mutagenesis may allow for development of these 14 phenotypes in a relatively small population of closely related plants, which should simplify 15 deduction of causal mutants by genomic resequencing. 16

2. Multiple disciplines. An interdisciplinary team is required to meet these objectives. In
 addition to colleagues in applied fields: plant pathologists, entomologists, agroecologists,
 agricultural economists, food or fuel chemists, colleagues from the biology departments--those
 studying evolution, plant physiology, plant-microbe ecology--are crucial if we are to design a
 new kind of domestication and document our efforts for future researchers. Engineers and
 computer scientists have already begun tackling some of the logistical and technical challenges
 arising from working with oddly shaped seed and large breeding populations of perennials.

Page 17 of 32

Electronic communication, online databases, and even social media make this kind of
 collaboration much easier than in the past.

3. Multiple breeding populations. Strong, recurrent artificial selection on replicate 3 small populations followed by hybridization to combine alleles from each selection regime has 4 5 been proposed as a strategy for accelerating domestication while minimizing genetic erosion 6 (DeHaan and Van Tassel, 2014). Natural selection acting on small, independently evolving 7 populations can also be exploited to obtain adapted (but not necessarily domesticated) biotypes. Marker assisted selection will be essential for rapidly eliminating linkage drag when 8 9 introgressing traits of interest from one population to another. More broadly, this combination of sequence informed background selection and the maintenance of numerous populations will 10 allow us to minimize the loss of diversity linked to selected sites which can occur via selective 11 sweeps. 12

After developing the *Silphium* composite cross base domestication population by crossing 13 14 genetically diverse accessions without selection (Allard, 1988); the seed will be distributed to collaborators in as many geographic locations as possible. Initially, indirect/natural selection will 15 be dominate at the sites, although some direct selection for reduced stratification requirements, 16 lodging resistance, and photoperiod responses may be required to retrieve seed each year from 17 the population. Seed will be harvested from each site every 2-4 years and replanted at the same 18 site. The small population should adapt to the local abiotic and biotic conditions, resulting in 19 20 gradual narrowing of genetic diversity at each site due to genetic drift and selection, but replication across several diverse sites should maintain a broad-genetic base in the crop as a 21 22 whole. The simplicity of this design enables the participation of researchers with a range of resources as well as farmers and non-research organizations. 23

1 4. Multiple environments. Selection for adaptation to multiple environments can ensure that future cultivars can be used across a broad landscape. Advanced materials distributed to 2 collaborators in multiple environments will also be used for G x E studies. Identifying traits 3 4 correlated with high performance in specific environments will be useful in further refining the ideotype. We expect that GWAS studies can quickly identify major domestication genes or at 5 least markers in linkage disequilibrium with the causal variants (Kruglyak, 2008). Even without 6 knowledge of causal genes / variants, knowledge of these genotype / phenotype (or 7 environmental) associations for Silphium domestication traits should lead to real applications in 8 the form of marker-assisted selection strategies which have been effectively used in many other 9 species, especially in the selection of simple monogenic traits and in pyramiding of major 10 disease resistance genes (Collard et al., 2008). 11

12

### 13 CONCLUSION

We are not yet ready to define precise ideotypes for Silphium oilseed, bioenergy or dual-14 purpose crops, but this thought-experiment has helped us identify areas of research needed in 15 order to do so (Figure 1). It also suggests to us that the ideotype concept itself needs to be 16 reinvented. In addition to the more familiar components relating to the classic domestication 17 syndrome and harvest index (Donald and Hamblin, 1983), we suggest that ideotypes for the next 18 generation of new crops need to identify how structural and functional plant features relate to 19 specific ecosystem services, what, if any, are the trade-offs involved between these characters, 20 21 what are the genetic bases for these traits, and thus, which selection method should be applied. New ideotypes should also include plans for retaining specific resource-use efficiency, stress-22

toleration and defensive adaptations during domestication. Achieving any of these goals using 1 breeding necessarily requires the fixation of major effect "domestication genes" at the plant level 2 and the loss of some genetic diversity from breeding populations, particularly in the case of 3 outcrossing perennials that likely harbor a high level of genetic load. However, the ideotype for 4 the quantity and structure of genetic diversity should extend to the whole crop through strategic 5 partitioning of diversity between populations under different types and levels of selection. 6 7 Finally, it is now possible to plan for what might be called meta-domestication: the data, narratives, analysis and biological samples that could allow a domestication event to be analyzed 8 and even replicated experimentally, in order to improve the ideotypes, increase the rate and 9 reduce the cost of future domestication projects. 10

11

12

REFERENCES

2	Allard, R.W. 1988. Genetic changes associated with the evolution of adaptedness in cultivated
3	plants and their wild progenitors. J. Hered. 79:225-238
4	Allendorf, F.W., P.A. Hohenlohe, and G. Luikart. 2010. Genomics and the future of conservation
5	genetics. Nat. Rev. Genet. 11:697-709.
6	Arkhipenko F.N., Larina V.I. 2011. Cup plant (Silphium perfoliatum L.) in the Ukranian forest
7	steppe. (In Russian.) Plant Introduction 1:9-13.
8	Asbjornsen, H. 2013. Targeting perennial vegetation in agricultural landscapes for enhancing
9	ecosystem services. Agric. Food Syst. 1:1–25.
10	Assefa, T. J. W, K.A. Albrecht, P.J. Johnson, and A. Boe. 2015. Genetic variation for biomass
11	and related morphological traits in cup plant (Silphium perfoliatum L.). Am. J. Plant Sci.
12	6:1098–1108.
13	Bai, C., W. S. Alverson, A. Follansbee, and D. M. Waller. 2012. New reports of nuclear DNA
14	content for 407 vascular plant taxa. Ann. Bot. 110:1623-1629.
15	Bare, J.E. 1979. Wildflowers and weeds of Kansas. The Regents Press of Kansas, Lawrence.
16	Baute, G. J., N.C. Kane, C.J. Grassa, Z. Lai, and L.H. Rieseberg. 2015. Genome scans reveal
17	candidate domestication and improvement genes in cultivated sunflower, as well as post-
18	domestication introgression with wild relatives. New Phytol. 206:830-838.
19	Benrey, B., A. Callejas, L. Rios, K. Oyama, and R.F. Denno. 1998. The effects of domestication
20	of Brassica and Phaseolus on the interaction between phytophagous insects and
21	parasitoids. Biol. Control 11:130-140.
22	Boeven, P.H., C.F.H. Longin, and T. Würschum. 2016. A unified framework for hybrid breeding
23	and the establishment of heterotic groups in wheat. Theor. Appl. Genet. 129:1231–1245.

1	Brown, A.H.D. 1989. The case for core collections. p. 136–156. In: Brown, A.D.H., D.R.
2	Marshall, O.H. Frankel, J.T. Williams (eds.). The use of plant genetic resources.
3	Cambridge University Press, Cambridge.
4	Calus, M.P.L. and R.F. Veerkamp. 2011. Accuracy of multi-trait genomic selection using
5	different methods. Genet. Select. Evol. 43:26 doi: 10.1186/1297-9686-43-26
6	Cameron, N.D. 1997. Selection indices and prediction of genetic merit in animal breeding. CAB
7	International. New York, NY.
8	Clark, L. V., J.E. Brummer, K. Glowacka, M.C. Hall, K. Heo, J. Peng, T. Yamada, J.H. Yoo,
9	C.Y. Yu, H. Zhao, S.P. Long, and E.J. Sacks. 2014. A footprint of past climate change on
10	the diversity and population structure of Miscanthus sinensis. Ann. Bot. 114:97–107.
11	Clark, L. V., J. Ryan Stewart, A. Nishiwaki, Y. Toma, J.B. Kjeldsen, U. J. Argensen, H. Zhao, J.
12	Peng, J.H. Yoo, K. Heo, C.Y. Yu, T. Yamada, and E.J. Sacks. 2015. Genetic structure of
13	Miscanthus sinensis and Miscanthus sacchariflorus in Japan indicates a gradient of
14	bidirectional but asymmetric introgression. J. Exp. Bot. 66:4213-4225.
15	Clevinger, J.A., and J.L. Panero. 2000. Phylogenetic analysis of Silphium and subtribe
16	Engelmanniinae (Asteraceae: Heliantheae) based on ITS and ETS sequence data. Am. J.
17	Bot. 87:565–572.
18	Collard, B.C.Y., D.J. Mackill, and P.T.R.S. B. 2008. Marker-assisted selection: an approach for
19	precision plant breeding in the twenty-first century. Phil. Trans. R. Soc. B 363:557-572.
20	Cowling, W.A., B.J. Buirchell, and D.E. Falk. 2009. A model for incorporating novel alleles
21	from the primary gene pool into elite crop breeding programs while reselecting major
22	genes for domestication or adaptation. Crop Pasture Sci. 60:1009–1015.
23	Crews, T.E., and L.R. DeHaan. 2015. The strong perennial vision: a response. Agroecol. Sustain.

1	Food Syst. 39:500–515
2	Crews, T.E., Blesh, J., Culman, S.W., Haye, R.C., Steen Jensen, E., Mack, M.C., Peoples, M.B.
3	and Schipanski, M.E. 2016. Going where no grains have gone before: from early to mid-
4	succession. Agriculture, Ecosystems and Environment 223:223-238
5	DeHaan, L.R., D.L. Van Tassel, J.A. Anderson, S.R. Asselin, R. Barnes, G.J. Baute, et al. 2016.
6	A pipeline strategy for grain crop domestication. Crop Sci. 56(3): 917–930.
7	DeHaan, L.R., and D.L. Van Tassel. 2014. Useful insights from evolutionary biology for
8	developing perennial grain crops. Am. J. Bot. 101:1801-19.
9	DeHaan, L.R., D.L. Van Tassel, and T.S. Cox. 2007. Perennial grain crops: a synthesis of
10	ecology and plant breeding. Renew. Agric. Food Syst. 20:5-14.
11	Desta, Z.A., and R. Ortiz. 2014. Genomic selection: Genome-wide prediction in plant
12	improvement. Trends Plant Sci. 19(9):592-601
13	Dhillion, S.S., and C.F. Friese. 1992. The occurrence of mycorrhizas in prairies: Application to
14	ecological restoration. In: The Proceedings of the 13th North American Prairie
15	Conference, University of Windsor (Canada). p.103-114.
16	Diamond, J. 2002. Evolution, consequences and future of plant and animal domestication. Nature
17	418:700–707.
18	Donald, C.M., and J. Hamblin. 1983. The convergent evolution of annual seed crops in
19	agriculture. Adv. Agron. 36:97-143.
20	Duvick, D.N. 1984. Genetic diversity in major farm crops on the farm and in reserve. Econ. Bot.
21	38:161–178.
22	Esquinas-Alcázar, J. 2005. Science and society: protecting crop genetic diversity for food
23	security: political, ethical and technical challenges. Nat. Rev. Genet. 6:946-953.

1	Ferchaud, F., G. Vitte, and B. Mary. 2016. Changes in soil carbon stocks under perennial and
2	annual bioenergy crops. GCB Bioenergy 8:290-306.
3	Fiedler, A.K., and D.A. Landis. 2007. Attractiveness of Michigan native plants to arthropod
4	natural enemies and herbivores. Environ. Entomol. 36:751-765.
5	Fontanesi, L., E. D'Alessandro, E. Scotti, L. Liotta, A. Crovetti, V. Chiofalo, and V. Russo.
6	Genetic heterogeneity and selection signature at the KIT gene in pigs showing different
7	coat colours and patterns. Animal Genetics 41:478-492.
8	Friesen, P.C., M. de Melo Peixoto, D.K. Lee, R.F. Sage. 2015. Sub-zero cold tolerance of
9	Spartina pectinata (prairie cordgrass) and Miscanthus × giganteus: candidate bioenergy
10	crops for cool temperate climates J. Exp. Bot. 66: 4403–4413.
11	Gansberger, M., L.F.R. Montgomery, and P. Liebhard. 2015. Botanical characteristics, crop
12	management and potential of Silphium perfoliatum L. as a renewable resource for biogas
13	production: A review. Ind. Crops Prod. 63:362-372.
14	Glover, J.D., J.P. Reganold, L.W. Bell, J. Borevitz, E.C. Brummer, E.S. Buckler, et al. 2010.
15	Increased food and ecosystem security via perennial grains. Science 328:1638–1639.
16	González-Paleo, L., and D.A. Ravetta. 2011. Indirect changes associated with a selection
17	program for increased seed-yield in wild species of Lesquerella (Brassicaceae): Are we
18	developing a phenotype opposite to the expected ideotype? Ind. Crops Prod. 34:1372-
19	1380.
20	González-Paleo, L., and D.A. Ravetta. 2012. Allocation patterns and phenology in wild and
21	selected accessions of annual and perennial Physaria (Lesquerella, Brassicaceae). Euph.
22	86:289–302.

23 González-Paleo, L., A.E Vilela, and D.A. Ravetta. 2016. Back to perennials. Does selection

1	enhance tradeoffs between yield and longevity? Ind. Crops. Prod. 91:272–278.
2	Han, K.J., K.A. Albrecht, R.E. Muck, and D.A. Kim. 2000a. Moisture effects on fermentation
3	characteristics of cup plant silage. Asian-Aus. J. Anim. Sci. 13:636-640.
4	Han, K.J., K.A. Albrecht, D.R. Mertens, and D.A. Kim. 2000b. Comparison of in vitro
5	digestion kinetics of cup plant and alfalfa. Asian-Aus. J. Anim. Sci. 13:641-644.
6	Harfouche, A., R. Meilan, M. Kirst, M. Morgante, W. Boerjan, M. Sabatti, et al. 2012.
7	Accelerating the domestication of forest trees in a changing world. Trends Plant Sci.
8	17:64–72.
9	Haussman, B.I.G., H.K. Parzies, T. Presterl, Z. Susic and T. Miedaner. 2004. Plant genetic
10	resources in crop improvement. Plant Genet Resour. 2:3-21.
11	Heffner, E.L., A.J. Lorenz, J.L. Jannink, M.E. Sorrells. 2010. Plant breeding with genomic
12	selection: gain per unit time and cost. Crop Sci. 50:1681–1690.
13	Henry, R.J. 2012. Next-generation sequencing for understanding and accelerating crop
14	domestication. Brief. Funct. Genomics 11:51–56.
15	Jamiolkowska, A., and R. Kowalski. 2012. In vitro estimate of influence of Silphium perfoliatum
16	L . leaves extract on some fungi colonizing the pepper plants. Agnieszka Jamiołkowska,
17	A, R Kowalski, R. Acta Sci. Pol. 11:43–55.
18	Jia, Y. and J.L. Jannink. 2012. Multiple-trait genomic selection methods increase genetic value
19	prediction accuracy. Genetics. 192:1513–1522.
20	Johnson, P., A. Boe, K. Albrecht, and V.C. Torrez. 2012. Recent discoveries and development in
21	the entomology of bioenergy crop production. Proceedings of the 2012 Sun Grant
22	National Conference. p. 4.
23	Juenger, T.E. 2013. Natural variation and genetic constraints on drought tolerance. Current

1	Opinion in Plant Biology 16:274–281
2	Kantar, M., C. Tyl, K. Dorn, X. Zhang, J. Jungers, J.M. Kaser, et al. 2016. Perennial grain and
3	oilseed crops. Annu. Rev. Plant Biol. 67: 703-729.
4	Kowalski, R., and J. Wiercinski. 2004. Evaluation of chemical composition of some Silphium L.
5	species seeds as alternative foodstuff raw materials. Pol. J. Food Nutr. Sci. 13:349-354.
6	Koziol, L., and J. Bever. 2015. Mycorrhizal response trades off with plant growth rate and
7	increases with plant successional status. Ecology 96:1768-1774.
8	Kruglyak, L. 2008. The road to genome-wide association studies. Nat. Rev. Genet. 9: 314-318.
9	Kula, R.R., P.J. Johnson, T.T. Heidel-Baker and A. Boe. 2017. A new species of Acanthocaudus
10	Smith (Braconidae: Aphidiinae), with a key to species and new host and distribution
11	records for aphidiines assolated with Silphium perfoliatum L. (Asterales: Asteraceae).
12	Zooaxa 4236(3) 543-552.
13	Ladizinsky, G. 1985. Founder effect in crop-plant evolution. Economic Botany. 39:191–199.
14	Lehmkuhler, J.W., M.H. Ramos, and K.A. Albrecht. 2007. Cup plant silage as a replacement for
15	corn silage in beef rations. Forage and Grazinglands. doi: 10.1094. FG-2007-1107-01-
16	RS, 2007.
17	Leopold, A. 1968. A Sand County almanac. 2 edition. Oxford University Press: Oxford.
18	
19	Lynch, J. P. 2013. Steep, cheap and deep: an ideotype to optimize water and N acquisition by
20	maize root systems. Ann. Bot. 112:347-357.
21	Mandel, J.R., S. Nambeesan, J.E. Bowers, L.F. Marek, D. Ebert, L.H. Rieseberg, S.J. Knapp,
22	and J.M. Burke. 2013. Association mapping and the genomic consequences of selection
23	in sunflower. Plos Genet. 9: e1003378. doi:10.1371/journal.pgen.1003378

1	Meilleur, B.A., and T. Hodgkin. 2004. In situ conservation of crop wild relatives : status and
2	trends. Biodivers. Conserv. 13:663-684.
3	National Plant Data Team, USDA-NRCS, 2016. The PLANTS Database. http://plants.usda.gov.
4	(accessed 20 Sep. 2016).
5	Ouyang, K., K. Wang, D. Li, Z. Li, and B. Liu. 2007. Establishment of sown pastures in the hilly
6	red soil region of the subtropics in southern China. Trop. Grasslands 41:92–99.
7	Pavek, D.S., W.F. Lamboy, and E.J. Garvey. 2003. Selecting in situ conservation sites for grape
8	genetic resources in the USA. Genet. Resour. Crop Evol. 50:165–173.
9	Pestsova, E. 2015. Silphium perfoliatum resource evaluation and development (SPREAD).
10	Bioeconomy Science Center. http://www.biosc.de/spread (accessed 28 September 2016).
11	Power, A.G. 2010. Ecosystem services and agriculture: tradeoffs and synergies. Philos. Trans. R.
12	Soc. Lond. B. Biol. Sci. 365:2959–2971.
13	Prasifka, J.R., D.K. Lee, J.D. Bradshaw, A.S. Parrish, and M.E. Gray. 2012. Seed reduction in
14	prairie cordgrass, spartina pectinata Link., by the floret-feeding caterpillar Aethes
15	spartinana (Barnes and McDunnough). BioEnergy Res. 5:189-196
16	Prezeworski, M., G. Coop, and J.D. Wall. 2005. The signature of positive selection on standing
17	genetic variation. Evolution. 59:2312–2323.
18	Raymer, P.L. 2002. Canola: An emerging oilseed crop. p. 122–126. In: Janick, J., and A.
19	Whipkey (eds.), Trends in new crops and new uses. ASHS Press, Alexandria, VA.
20	Resende, M.D.V., M.F.R. Resende Jr., C.P. Sansaloni, C.D. Petroli, A.A. Missiaggia, A.M.
21	Aguiar, et al. 2012. Genomic selection for growth and wood quality in <i>Eucalyptus</i> :
22	capturing the missing heritability and accelerating breeding for complex traits in forest
23	trees. New Phytol. 194:116–128.

1	Rellstab, C., F. Gugerli, A. J. Eckert, A. M. Hancock and R. Holderegger 2015. A practical guide
2	to environmental association analysis in landscape genomics. Mol. Ecol. 24: 4348-4370.
3	Reyno, R., R. Narancio, P. Speranza, J. Do Canto, B. López-Carro, P. Hernández, J. Burgueño,
4	D. Real, and M.D. Rizza. 2012. Molecular and cytogenetic characterization of a
5	collection of bahiagrass (Paspalum notatum Flügge) native to Uruguay. Genet. Resour.
6	Crop Evol. 59:1823–1832.
7	Rodriguez-Saona, C., N. Vorsa, A.P. Singh, J. Johnson-Cicalese, Z. Szendrei, M.C. Mescher,
8	and C.J. Frost. 2011. Tracing the history of plant traits under domestication in
9	cranberries: potential consequences on anti-herbivore defenses. J. Exp. Bot. 62: 2633-44.
10	Rozema, J., and T. Flowers. 2008. Crops for a salinized world. Science. 322:1478–1480.
11	Runck, B.C., M.B. Kantar, N.R. Jordan, J.A. Anderson, D.L. Wyse, J.O. Eckberg, et al. 2014.
12	The reflective plant breeding paradigm: A robust system of germplasm development to
13	support strategic diversification of agroecosystems. Crop Sci. 54:1939–1948.
14	Sanderson, M.A., and P.R. Adler. 2008. Perennial forages as second generation bioenergy crops.
15	Int. J. Mol. Sci. 9:768–788.
16	Saxena, K.B., and S.L. Sawargaonkar. 2014. First information on heterotic groups in pigeonpea
17	[Cajanus cajan (L.) Millsp.]. Euphytica 200:187–196
18	
19	Schoo, B., K.P. Wittich, U. Böttcher, H. Kage, and S. Schittenhelm. 2016. Drought tolerance and
20	water-use efficiency of biogas crops: a comparison of cup plant, maize and lucerne-grass.
21	J. Agron. Crop Sci. doi.10.1111/jac.12173
22	Schorpp, Q., and S. Schrader. 2016. Earthworm functional groups respond to the perennial
23	energy cropping system of the cup plant (Silphium perfoliatum L.). Biomass and

1	Bioenergy 87:61–68.	
2	Seifert, E.K., J.D. Bever, and J.L. Maron. 2009. Evidence for the evolution of reduced	
3	mycorrhizal dependence during plant invasion. Ecology 90:1055–1062.	
4	Settle, W.J. 1967. The Chromosome Morphology in the Genus Silphium (Compositae). Ohio J.	
5	Sci. 67:10–19.	
6	Shapter, F.M., M. Cross, G. Ablett, S. Malory, I.H. Chivers, G.J. King, and R.J. Henry. 2013.	
7	High-throughput sequencing and mutagenesis to accelerate the domestication of	
8	Microlaena stipoides as a new food crop. PLoS One 8: e82641	
9	doi:10.1371/journal.pone.0082641	
10	Sharma, H.C., Crouch, J.H., Sharma, K.K., Seetharama, N., Hash, C.T. 2002. Applications of	
11	biotechnology for crop improvement: prospects and constraints. Plant Science 163:381-	
12	395	
13	Simmonds, N.W. 1962. Variability in crop plants, its use and conservation. Biol. Rev. 37:422-	
14	465.	
15	5 Simmonds, N.W. 1993. Introgression and incorporation - strategies for the use of crop genetic	
16	resources. Biol. Rev. Camb. Philos. Soc. 68:539-562.	
17	Siol, M., S.I. Wright, and S.C.H. Barrett. 2010. The population genomics of plant adaptation.	
18	New Phytol. 188:313–332	
19	Smaje, C. 2015. The strong perennial vision: a critical review. Agroecol. Sustain. Food Syst.	
20	39:471–499	
21	Stanford, G. 1990. Silphium perfoliatum (cup-plant) as a new forage. In: Proceedings of the	
22	twelfth North American prairie conference. University of Northern Iowa, Cedar Falls. p.	
23	33–37	

1	The Plant List. 2013. Version 1.1. http://www.theplantlist.org (accessed 10 Sep. 2016).			
2	Tuell, J.K., A.K. Fiedler, D. Landis, and R. Isaacs. 2008. Visitation by wild and managed bees			
3	(Hymenoptera: Apoidea) to Eastern U . S . native plants for use in conservation			
4	programs. Environ. Entomol. 37:707-718			
5	Van Tassel, D.L., S.R. Asselin, S.A. Cox, G. Sideli, and D.J. Cattani. 2014. Evaluating perennia			
6	candidates for domestication: lessons from wild sunflower relatives. In: Batello, C., L.			
7	Wade, S. Cox, N. Pogna, A. Bozzini, J. Choptiany. (eds.). Perennial crops for food			
8	security; Proceedings of the FAO Expert Workshop. Rome. p. 112-140.			
9	Van Tassel, D.L., L.R. DeHaan, and T.S. Cox. 2010. Missing domesticated plant forms: can			
10	artificial selection fill the gap? Evol. Appl. 3:434-452.			
11	Vico, G., S. Manzoni, K. Murphy, and M. Weih. 2015. Trade-offs between seed output and life			
12	span – a quantitative comparison of traits between annual and perennial congeneric			
13	species. New. Phytol. 209:204-2014			
14	Visscher, P. M., M. A. Brown, M. I. McCarthy and J Yang. 2012. Five Years of GWAS			
15	Discovery. Am. J. Hum. Genet. 90:7–24.			
16	Von Gehren, P., M. Gansberger, J. Mayr, and P. Liebhard. 2016. The effect of sowing date and			
17	seed pre-treatments on establishment of the energy plant Silphium perfoliatum L. by			
18	sowing. Seed Sci. Tech. 44:1–10.			
19	Weaver, J.E., L. a. Stoddart, and W. Noll. 1935. Response of the prairie to the great drought of			
20	1934. Ecology 16:612–629.			
21	Wright, S.I., I.V. Bi, S.G. Schroeder, M. Yamasaki, J.F. Doebley, M.D. McMullen, and B.S.			
22	Gaut. 2005. The effects of artificial selection on the maize genome. Science 308:1310-			
23	1314.			

1	Wrobel, M., J. Fraczek, S. Francik, Z. Slipek, and M. Krzysztof. 2013. Influence of degree of		
2	fragmentation on chosen quality parameters of briquette made from biomass of cup plant		
3	Silphium perfoliatum 1. Jelgava 23:653–657.		
4	Wynia, R. 2009. Plant Guide for compass plant (Silphium laciniatum L.). USDA-Natural		
5	Resources Conservation Service, Kansas Plant Materials Center, Manhattan, Kansas.		
6	Yatskievych, G. 2006. Steyermark's Flora of Missouri.Volume 2. Revised ed. Missouri		
7	Botanical Garden Press, St. Louis, Missouri.		
8	Zhang, H., Z. Shen, G. Yang, Z. An, and Q. Sun. 2011. Effect of chicken manure-amended		
9	copper mine tailings on growth of compositae Silphium perfoliatum and substrate		
10	properties. Southwest China J. Agric. Sci. 24:1792-1798.		
11	Zhang, Q., Q. Zhou, L. Ren, Y. Zhu, and S. Sun. 2006. Ecological effects of crude oil residues		
12	on the functional diversity of soil microorganisms in three weed rhizospheres. J. Environ.		
13	Sci. 18:1101–1106.		
14 15			

#### 1 **Figure Legends**

2

3 Figure 1. Hypothetical stages of *de novo* domestication of *Silphium* as an oilseed grain. To

avoid developing new crops that are as functionally deficient in ecosystems as our current, 4

5 annual food and energy crops, recent domestication proposals focus on species with adaptations

6 quite distinct from the ancestors of existing crops (e.g., perennials, halophytes). Rapid

improvement of wild candidates (left column) for key domestication traits is required to make 7

these new crops economically viable. Under the hypothetical scenario depicted here, "over-8

9 domestication" could occur if strong directional selection for yield traits resulted in the loss of

desirable adaptations present in the wild progenitor (right column), through genetic drift or 10

- correlated responses to selection. An ideotype designed to deliver a balanced range of crop 11
- services (center column) will be a complex mixture of traits: some nearly indistinguishable from 12 annual crops (yellow) and other very close to the fully "wild" state (green). A third set of traits
- 13 will likely be intermediate between fully wild and fully domesticated (green and yellow) either
- 14 because a desirable adaptation is "overbuilt" for survival and vigor in a managed ecosystem or 15
- because the time and cost required to fully eliminate the wild trait is not justified by the potential 16
- benefit. Blended transitions between yellow and green represent our present uncertainty; 17
- multidisciplinary collaboration will be required to identify the costs and benefits of modifying
- 18
- 19 each trait.

20

Undomesticated ecotype —		> "Over-domesticated" cultivar
		Cuitivai
Low and/or variable whole-plant harvest index	Above-ground harvest index may approach that of annual crops but large reduction in belowground biomass could reduce survival and ecosystem services.	High and consistent whole-plant harvest index
Very long lifespan	Intermediate lifespan may provide acceptable services and stand replacement may be necessary for long-term crop rotation and to introduce improved cultivars.	Annual or short-lived perennial
Investment in traits to acquire resources unavailable to most crops	Diminishing returns on investment may justify moderated acquisition e.g., of water from extreme depth, photoenergy very late in the season, etc.	Adapted to acquiring resources from a more limited time and space
Adaptation to intraspecific competition	Intermediate reduction in plant height, spreading roots, etc., allows increased yield per unit area without excessively reducing competitiveness with weeds.	Intraspecific competition minimized
Many heads, numerous and complex inflorescences, long flowering period	High yield could be achieved with multiple heads per stem but a reduction in the number of heads may be necessary to achieve synchrony.	Determinate, single-head, unbranched, highly synchronized flowering
Seeds and heads adapted for efficient dispersal	Humans must take over the function of seed dispersal for efficient harvesting.	Completely non-shattering heads. Loss of dispersal structures
Grain not suitable for processing with standard equipment	Difficulties in harvesting, cleaning, processing the grain may prevent profitable adoption of the crop.	Grains easy to process and use commercially
Seed dormancy prevents germination in unfavorable conditions	Farmers can predict/create safe germination conditions. Rapid, synchronous germination facilitates weed control and a uniform stand.	Rapid germination and stand establishment
Ability to recover from severe damage by rapid reallocation of reserves	Intermediate resilience justified because frequency of disturbances is lower in managed landscapes.	Poor resilience to sudden stress due to low reserves and reduced plasticity.
Physiological and morphological adaptation for abiotic stress tolerance	Maintaining resistance to stresses is desirable for reducing inputs but some adaptations may be unnecessary in a human-managed environment.	Many abiotic stresses relaxed using expensive or non-renewable inputs
Enemies limited by polygenic defense mechanisms and resistance genes	Disease and pest pressures may be higher in large, high density agricultural stands than in the wild, so preservation of defense and resistance genes within the primary gene pool should be prioritized.	Highly reliant on pesticides to limit enemies. Wide-crossing required to acquire resistance genes.
Smaller, better defended leaves with high nitrogen and water use efficiency	Both increased seedling growth rate and retention of resource use efficiency are desirable.	Bigger, thinner leaves with reduced construction costs, rapid growth
Highly dependent upont mutualists for nutrients	Avoiding breeding in excessively fertile environments can help maintain efficient mutualisms but fertilization at specific developmental stages may be needed for adequate yield.	Dependent upon inputs and/or tillage for nutrients
Pollen and pollinator rewards abundant	Supporting pollinators is an increasingly valued agro- ecological service but achieving other domestication goals may require compromise in floral traits.	Few floral resources for pollinators
Seeds, biomass rich in secondary metabolites with industrial, flavor, or nutritional value	Selection for increased seed oil, starch, and protein may reduce the concentration of other phytochemicals unless they are identified as desirable and selected for.	Low levels of secondary metabolites in grain and/or biomass
High genetic diversity	Relatively few domestication alleles must be fixed but breeding strategies can minimize unintended allele loss through hitchhiking or drift.	Low genetic diversity in primary gene pool